CAMBRIDGE INTERNATIONAL EXAMINATIONS

Cambridge International Advanced Subsidiary and Advanced Level

MARK SCHEME for the May/June 2015 series

9700 BIOLOGY

9700/52

Paper 5 (Planning, Analysis and Evaluation), maximum raw mark 30

This mark scheme is published as an aid to teachers and candidates, to indicate the requirements of the examination. It shows the basis on which Examiners were instructed to award marks. It does not indicate the details of the discussions that took place at an Examiners' meeting before marking began, which would have considered the acceptability of alternative answers.

Mark schemes should be read in conjunction with the question paper and the Principal Examiner Report for Teachers.

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Mark scheme abbreviations:

; / separates marking points

alternatives answers for the same point

R reject

Α accept (for answers correctly cued by the question, or extra guidance)

AW alternative wording (where responses vary more than usual)

<u>underline</u> actual word given must be used by candidate (grammatical variants accepted)

max indicates the maximum number of marks that can be given

or reverse argument ora error carried forward ecf

ignore

mp marking point (with relevant number)

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Question	Expected answer	Extra guidance	Mark
1 (a)	independent: concentration of amylase/enzyme;	I amount R (amylase) extract	
	dependent: diameter / area of brown zone;	 A radius of brown zone A starch free/digested starch/clear zone A area of brown zone minus well R blue zone 	[2]
(b)	any 8 from: independent variable 1 ref. to method of diluting the 0.5 g dm ³ amylase/stock amylase solution and to give a minimum of 5 dilutions;	 0.0 and 0.5 g dm³ can be included in the number of dilutions A serial/series/simple/proportional/dilution as method OR a description. Use the formula C₁V₁ = C₂V₂ to make A If the fungal extract is diluted instead of amylase but R mp5 	
	2 ref. to concentrations from 0.5 g dm ³ downwards with correct units;	2 minimum of 3 other stated values between 0.5 g dm ³ and 0.0 g dm ³ must correspond to dilution method chosen ecf if no method given A 10 fold or a 50% reduction serial dilution	
	3 use of a <u>control</u> with example ;		
	dependent variable 4 ref. to a suitable method of measuring diameters/width/	3 A water/0.0 g dm ³ /boiled or denatured extract/enzyme	
	radius/area of brown zones ;	 4 e.g. using (suitable) ruler/callipers/string and ruler R metre ruler I graticules OR use a (transparent) grid/graph paper and count the number of squares OR take a photograph and measure using one the methods for finding area NOTE: method of measuring must match what they have stated they are measuring. 	

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5	ref. to testing the (fungal) extract/fungal amylase/fungal enzyme;	5	R if dilute the fungal extract	
6	plot a calibration curve of known concentrations and use it to determine extract concentration;	6	A compare extract result with results of known concentrations to identify extract concentration OR find approx. range of concentration and then do more at smaller intervals to identify extract	
sta	andardising variables (max. 3)		concentration	
7	ref. to suitable stated volume/same volume of amylase (in each well);	7	If a volume is stated – max 1 cm ³ I amount/known	
8	leave (all plates) for same period of time;	8	if time stated, minimum 30 min/max 24 hours	
9	method of maintaining at same/constant/optimum/stated temperature;	9	e.g. incubat <u>or</u> /constant temp. room A water bath if temp. stated, any single temp. in range 15–65°C I air conditioning	
10	use a buffer to keep the pH of the agar same ;	10	e.g. making or adding buffer to agar or starch (solutions) A adding buffer to the amylase solution before using it A if stated, any single pH	
11	same concentration of starch (in the agar plates);	11	<u> </u>	
12	same denth/volume of agar in Petri dish ·	12		
	·	12	I mass of agar/depth of wells	
sa	fety			
14	ref. to low risk investigation/hazard <u>and</u> suitable safety precaution;	14	fungal/enzyme allergy or fungus/enzyme/iodine/agar is irritant <u>and</u> wearing gloves/eye protection/mask I iodine as an allergen R no risk	
15	ref. to a minimum of three replicates <u>and</u> calculate a mean or identify/eliminate/remove anomalies;	15	A original and 2 more/several/many/multipleA outliers for anomaliesR reduce anomalies	[max 8
	6 sta 7 8 9 10 11 12 13 sa 14 rel	enzyme; 6 plot a calibration curve of known concentrations and use it to determine extract concentration; standardising variables (max. 3) 7 ref. to suitable stated volume/same volume of amylase (in each well); 8 leave (all plates) for same period of time; 9 method of maintaining at same/constant/optimum/stated temperature; 10 use a buffer to keep the pH of the agar same; 11 same concentration of starch (in the agar plates); 12 same depth/volume of agar in Petri dish; 13 cover to prevent contamination / evaporation; safety 14 ref. to low risk investigation/hazard and suitable safety precaution; reliability 15 ref. to a minimum of three replicates and calculate a mean or	enzyme; 6 plot a calibration curve of known concentrations and use it to determine extract concentration; standardising variables (max. 3) 7 ref. to suitable stated volume/same volume of amylase (in each well); 8 leave (all plates) for same period of time; 9 method of maintaining at same/constant/optimum/stated temperature; 10 use a buffer to keep the pH of the agar same; 11 same concentration of starch (in the agar plates); 12 same depth/volume of agar in Petri dish; 13 cover to prevent contamination / evaporation; safety 14 ref. to low risk investigation/hazard and suitable safety precaution; reliability 15 ref. to a minimum of three replicates and calculate a mean or	enzyme; 6 plot a calibration curve of known concentrations and use it to determine extract concentration; 8 standardising variables (max. 3) 7 ref. to suitable stated volume/same volume of amylase (in each well); 8 leave (all plates) for same period of time; 9 method of maintaining at same/constant/optimum/stated temperature; 10 use a buffer to keep the pH of the agar same; 10 use a buffer to keep the pH of the agar same; 11 same concentration of starch (in the agar plates); 12 same depth/volume of agar in Petri dish; 13 cover to prevent contamination / evaporation; 14 ferf. to low risk investigation/hazard and suitable safety precaution; 15 ref. to a minimum of three replicates and calculate a mean or 16 A compare extract result with results of known concentration on concentration on one family and use it to concentration on one family find and use it to concentration. 16 A compare extract result with results of known concentration on one family and use it to dentify extract concentration. 16 A compare extract result with results of known concentration on one family and use it to dentify extract concentration. 16 A compare extract result with results of known concentration on one family and usering one concentration. 16 A compare extract result with results of known concentration on one family and usering one at smaller intervals to identify extract concentration. 18 ref. to authorise stated — max 1 cm³ 1 amount/maw 24 hours 9 e.g. incubator/constant temp. room A water bath if temp. stated, any single temp. in range 15–65°C I air conditioning 10 e.g. making or adding buffer to agar or starch (solutions) A adding buffer to the amylase solution before using it A if stated, any single pH 11 I ref. to other nutrients in agar, I amount 12 A depth of agar plate I mass of agar/depth of wells 14 fungal/enzyme allergy or fungus/enzyme/iodine/agar is irritant and wearing gloves/eye protection/mask I iodine as an allergen R no risk

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(c)	use a (glucose) biosensor/glucose dipstick or test-strip/named test strip, e.g. clinistix/uristix;	A use Benedict's test and EITHER weigh precipitate OR use colorimeter to find the intensity of blue solution left OR time the first appearance of a colour change A chromatography and stain (to show sugars) A Barfoed's test for monosaccharides	
	(because) γ -amylase will produce glucose only / β -amylase will produce maltose (mainly) ;	I idea that γ -amylase produces more glucose than β-amylase R ora	[2]
(d) (i)	1963 shown on extract B , plate 5;		
	1809 shown on extract D , plate 3;		[2]
(ii)	reject/eliminate/ignore/leave out (affected data from calculations);	A repeat until consistent results obtained I repeat unqualified OR repeat to find a mean	[1]

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(iii)	correct values for $\Sigma x = 1820$					
	and $\bar{x} = 303/303.3/3$	303.3°			e.g. $\begin{array}{ c c c c c c c c c c c c c c c c c c c$	
	and $(x \overline{x})^2$;					
	$(x \ \overline{x})^2$ allowed 341	341.34	341.4 3	342		
	$(x \ \overline{x})^2$ not allowed 341.32	341.2			allow ecf for s from wrong value of $\sum (x \ \bar{x})^2$	
	correct value of s from table ;					
	correct values of s (±) 8.(0)	8.26 8.27	8.3			[max 2]
(iv)	any 3 from: 1 the larger/AW the brown area the more amylase/more enzyme activity ora			e enzyme	Ignore ref. to proportionality / positive correlation between enzyme concentration and gene copies	
	OR the larger/AW the brown a 2 (person or extract) A had a mylase/(person or extract amylase; 3 (person) A has the highes has the lowest number of	the high <u>est</u> /AW <u>c</u> ct) F has low <u>est</u> A	oncentration W concentr	<u>n</u> of <u>ration</u> of	A strongest/largest/most/ ora weakest/smallest/least A A>B>C>D>E>F I copying results without any conclusion about rank order	
	4 ref. to genetic variation in amylase);	population (for the	production	n of		[3]
					Total:	[20]

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2	(a)	any 3 from: variation in volunteers body mass/weight;	Mark the first three given. I environmental factors, e.g. temperature light noise in room time of day I diet	
		2 ref. to use of drugs e.g. medication/self-inflicted;	I ref. to distance between or position of electrodes/time of charge application	
		3 alcohol consumption ;		
		4 smoking status ;		
		5 ref. to ethnicity;		
		6 volunteer all have the same 'handedness' ;		
		7 no medical/named medical condition affecting nerve conduction ;	7 e.g. Multiple Sclerosis(MS), Myalgic Encephalitis(ME), Muscular Dystrophy(MD), Motor Neurone Disease (MND),spinal bifida, polio A disease of the nervous system	
		variation in method of applying test 8 same arm tested ;		
		9 idea of volunteer not moving (during the test);	9 e.g. at rest/sitting/lying down	
		10 electrical charge/coulombs/voltage/potential difference;	10 A electrical stimulus I current	
		11 same number of volunteers in each age category;		[max 3]

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(b)	age/years	mean conduction velocity ± S _M	confidence limits		note: must have both numbers correct for one mark	
			lower limit	upper limit		[1]
	60–69	52.2 ± 0.675	<u>50.85</u>	<u>53.55</u> ;		
(c) (i)	30–39 and 70–79 OR 40–49 and 70–79 OR 50–59 and 70–79;				A descriptions, e.g. upper limit of 70–79 does not reach lower limit of 30–39 I error bars overlapping/range(bars) not overlapping I ref. to mean conduction velocity	
	there is no overlap of (confidence) limits/ S_M ;					[2]
(ii)	test: t test;				R continuous variation	
	reason: comparing (two) means/normal distribution/continuous data;					[2]
(iii)	there is no significant difference between the (mean) conduction velocities / NCV of (individuals from) different age groups;				needs to be clear that the significant difference is in the conduction velocity and the not the ages A the difference in (mean) conduction velocities of individuals from different age groups is not significant A stated age categories/women of different ages/young(er)	
					and old(er) women/with age	1
(d)	large sample size /number of people tested ;				A tested 394 individuals/many people/lots of people; I sufficient/enough people tested I number of different age categories/ref. to standard error	[1]
					Total:	[10]